



For the Life Scientist

HiFi-App Polymerase

CONCENTRATION: 2U/ μ l

Store at -20°C. (The kit will retain full activity for 12 months at -20°C. Can be stored at 4°C for 1 month and go through 30 freeze/thaw cycles with no loss of activity. Avoid prolonged exposure to light).

DESCRIPTION

HiFi-App Polymerase is a superior performance proofreading enzyme which has been specifically engineered for high fidelity PCR amplification of DNA fragments up to 10 kb. It has a 50 fold higher fidelity than AppTaq polymerase with an error rate of 1 error per 4.5 x 10⁷ nucleotides incorporated. It comes with a 5x HiFi reaction buffer which has pre-added enhancers, stabilisers, MgCl₂ and dNTPs to maximise PCR yields, and works in fast or standard thermal cycling conditions. The enzyme generates blunt ends on the PCR products which can then be used in blunt end cloning and site-directed mutagenesis.

ORDERING INFORMATION

| Component | ARP041 | ARP043 |
|-----------------------------------|----------------------------|-----------------------------|
| HiFi-App Polymerase (2U/ μ l) | 200 units (1x 100 μ l) | 1000 units (5x 100 μ l) |
| 5x HiFi reaction buffer | 3x 1ml | 15 x 1ml |

PROTOCOL

Prepare a PCR master mix by mixing molecular biology grade water, 5x HiFi reaction buffer, forward and reverse primers and HiFi-App Polymerase. Prepare sufficient master mix for the number of reactions plus one extra. Aliquot the master mix into individual PCR tubes / wells and then add template DNA.

1. Gently mix and briefly centrifuge all solutions after thawing.
2. Add the following components for each 50 μ L reaction to a thin-walled PCR tube/plate:

| Reagent | Final Concentration | 50 μ L reaction |
|---|---------------------|-------------------------------|
| 5x HiFi reaction buffer | 1X | 10.0 μ L |
| Forward primer (10 μ M) | 400nM | 2.0 μ L |
| Reverse primer (10 μ M) | 400nM | 2.0 μ L |
| Template DNA | 100 - 500ng* | variable |
| HiFi-App Polymerase (2U/ μ L) | 1.25U - 5U | 0.25 μ L – 1 μ L |
| Molecular Biology Grade water, (BMW001) | | Up to 50 μ l final volume |

3. Gently mix the samples and spin down.
4. If using a thermal cycler that does not use a heated lid, overlay the reaction mixture with 25 μ L of mineral oil.



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5. Perform PCR using recommended thermal cycling conditions:

| Step | Temperature / °C | Time | Number of cycles |
|----------------------|------------------|-----------|------------------|
| Initial denaturation | 95 | 1 min | 1 |
| Denaturation | 95 | 15 s | 25-40 |
| Annealing | 55-65 | 15 s | |
| Extension | 72 | 30 s / kb | |

CONSIDERATIONS

Template DNA*

For optimal results, use between 5ng and 500ng per reaction for eukaryotic DNA, and for cDNA use below 100ng in the 50µL reaction volume. Higher amount of template increases the risk of non-specific PCR products. Trace amounts of certain agents used for DNA purification, such as phenol, EDTA and proteinase K, can inhibit DNA polymerases. Ethanol precipitation and repeated washes of the DNA pellet with 70% ethanol normally removes trace contaminants from DNA samples.

Reaction Buffer

The 5x HiFi reaction buffer contains optimal concentrations of MgCl₂ (15mM) and dNTPs (5mM), as well as enhancers and stabilisers. This avoids having to vary these parameters to obtain maximum PCR yields. We do not recommend adding further enhancers or magnesium.

Primers

The recommended concentration range of the PCR primers is 0.1-1 µM. Excessive primer concentrations increase the probability of mis-priming and non-specific PCR products.

Denaturation

Complete initial denaturation of the template DNA is essential for efficient utilization of the template during the first amplification cycle. If the GC content of the template is 50% or less, an initial 1-3 min denaturation at 95°C is sufficient. For colony PCR, denature for 10 mins.

Annealing

The optimal annealing temperature is 5°C lower than the melting temperature (T_m) of the primers. Incubation for 0.25-2 min is usually sufficient. However, if non-specific PCR products are obtained in addition to the expected product, the annealing temperature should be optimized by increasing it stepwise by 1-2°C.

Extension

The optimal extension temperature for HiFi-App Polymerase is 72°C. The recommended extension time is 30 s per kb for amplification from eukaryotic DNA or cDNA.

TROUBLE SHOOTING / TECHNICAL SUPPORT

For troubleshooting please visit

www.appletonwoods.co.uk/PCRtroubleshooting.pdf for a trouble shooting guide on PCR. If this does not resolve your issues, please email technicalsupport@appletonwoods.co.uk with details of your: amplicon size, reaction setup, cycling conditions, gel images.

Note: HiFi-App Polymerase is for research use only.

ASSOCIATED PRODUCTS

| Product | Pack Size | Product Code |
|---------------------------------|-------------------|--------------|
| Molecular Biology Grade Agarose | 100g | AG002 |
| Molecular Biology Grade Agarose | 500g | AG001 |
| AxyPrep Mag PCR clean up Kit | 5mL, 110 preps | AX401 |
| AxyPrep Mag PCR clean up Kit | 50mL, 1110 preps | AX402 |
| AxyPrep Mag PCR clean up Kit | 250mL, 5550 preps | AX403 |
| Molecular biology grade water | 100mL | BMW001 |
| Molecular biology grade water | 500mL | BMW002 |

More pack sizes available at www.appletonwoods.co.uk